

## Abiotic Stress Responses in Plants: Potential Targets on Studying Heavy Metal Stress tolerance in Bryophytes

Rekha Tyagi<sup>1</sup>, Anjana Singh<sup>2\*</sup>, Anjuli Sood<sup>2</sup> and P.L. Uniyal<sup>2</sup>

<sup>1</sup>GGS IP University, Dwarka, Sector 16C, New Delhi-110078, INDIA; <sup>2</sup>Department of Botany, University of Delhi, Delhi-110007, INDIA

### Publication Info

#### Article history:

Received : 30.11.2016

Accepted : 15.06.2017

DOI : 10.18811/ijpen.v3i.8446

#### Key words:

Cell to cell communication

Mosses

Stress physiology

Stress response

#### \*Corresponding author:

Dr. Anjana Singh

Tel.: 09871157541

Email: anjana8@gmail.com

### Abstract

Pollution of the biosphere with heavy metals has phenomenally increased since the commencement of industrial revolution. It poses several environment and health concerns. High regeneration and metal accumulation capacity the possibility of genetic transformation by homologous recombination extend the candidature of bryophytes as promising experimental models for heavy metal stress tolerance studies. Plants use several abiotic stress pathways which share common elements that are potential "nodes" for cross talks. Common elements/ molecules, which are likely to occur early in several stress response cascades in bryophytes seem to be the potential targets for heavy metal tolerance studies that can be worked out in future, at biochemical, protein and gene level.

### 1. Introduction

Heavy metals are defined as metals with a density greater than 5 g cm<sup>-3</sup>. Most of the heavy metals are transition metals with an incompletely filled d-orbital, present as cations under physiological conditions. Fe, Mo, Cu, Zn and Mn are biologically significant, while As, Hg, Ag, Sb, Cd and Pb have no known function as nutrients and seem to be more or less toxic to plants (Goldbold and Huttermann, 1985; Breckle, 1991; Nies, 1999). The two main reasons for heavy metal's presence in atmosphere are natural sources and anthropogenic activities. Natural sources include weathering of bedrocks, volcanoes and continental dusts. Anthropogenic activities like mining, combustion of fossil fuels, phosphate fertilizers, etc. lead to the emission and accumulation of heavy metals in ecosystem (Lanzty and Mackenzie, 1979; Galloway *et al.*, 1982; Angelone and Bini, 1992).

Heavy metals can have numerous harmful effects on organisms. The ions of some of these heavy metals readily form ligands with organic molecules and tear them. Heavy metals affect enzyme functions, cell membrane permeability and cell growth (Moriarty, 1999). Metal pollutants disturb the species, community, and ecosystem as a whole through food chain (Nriagu, 1996). Toxic effects of heavy metals are exerted at the extracellular as well as intracellular levels in bryophytes. Two different uptake routes have been

identified: a) passive uptake, only driven by the concentration gradient across the membrane and b) active energy dependent uptake (Nies, 1999; Williams *et al.*, 2000).

Heavy metals can be divided into two groups of redox active and redox inactive metals. Autoxidation of redox active metals such as Fe<sup>2+</sup> or Cu<sup>+</sup> results in O<sub>2</sub><sup>-</sup> formation and subsequently in H<sub>2</sub>O<sub>2</sub> and OH<sup>-</sup> production via Fenton-type reactions. Cellular injury by this mechanism is reported for due to heavy metals (Jones *et al.*, 1991; Lund *et al.*, 1991; Shi *et al.*, 1993; Imlay *et al.*, 1998). Another important mechanism of heavy metal toxicity is its ability to bind strongly to oxygen, nitrogen and sulphur atoms. Direct effect of cadmium on the sulphhydryl homeostasis of cells and inhibition of enzymes has been reported for mammalian and animal cells (Chrestensen *et al.*, 2000). Heavy metals also harm by displacing the essential metal ions from enzymes. Divalent cations such as Co<sup>2+</sup>, Ni<sup>2+</sup> and Zn<sup>2+</sup> were found to displace Mg<sup>2+</sup> in ribulose-1, 5-bisphosphate-carboxylase/oxygenase and result in loss of activity (Van Assche and Clijsters, 1986; Rivetta *et al.*, 1997).

Plants have an inbuilt capacity to adjust to a variety of abiotic stress conditions. Abiotic stresses elicit complex cellular and molecular responses in plants that enable them to tolerate and survive adverse conditions. Plant's acclimatization to abiotic stresses is accompanied by a cascade of events which starts with

stress perception and ends with the expression of a set of target genes eventually leading to a variety of morphological, biochemical and physiological changes. Plants use common pathways and components in a stress response relationship. This phenomenon is called cross-tolerance whereby a plant adjusts to a range of different stresses after being exposed to one specific stress. Separate abiotic stress signaling pathways are likely to interact in a similar manner i.e. "Cross talks" can occur between different sensors and signal transduction pathways. It could be any instance of two signaling pathways from different stressors that converge (Knight and Knight, 2001). Several abiotic stress pathways share common elements that are potential "nodes" for cross talks. Factors that act at early stages of a stress response cascade are critical for other cell functions and significantly affect the operation of other genes. There are common signals/elements, which are likely to occur "early in the stress response cascades" (Pastori and Foyer, 2002). During stress they show higher superoxide dismutase (SOD) activity or lower activities of the chloroplastic  $H_2O_2$ -processing enzymes peroxidase and ascorbate peroxidase. Significant decreases in the activity of catalase and depletion of the anti-oxidant ascorbic acid is also reported. The role of activated oxygen-processing enzymes involved in the removal of chloroplastic hydrogen peroxide may be less important than that of anti-oxidants in the determination of stress tolerance (Oliver *et al.*, 2005).

## 2. Cross talks between different pathways can be mediated at different levels

### 2.1. Elevation in cytosolic free calcium levels

Elevation in  $[Ca^{2+}]_{cyt}$  is thought to be the primary stimulus sensing event in response to different environmental stresses (Sanders *et al.*, 1999; Knight 2000). Plants exhibit markedly different levels of elevation in  $[Ca^{2+}]_{cyt}$  when already exposed to a previous stress indicating a cross talk between abiotic stress signal transduction pathways occurring at the level of calcium (Knight *et al.*, 1998; Knight, 2000). In *Arabidopsis*, an oxidative stress encounter nullified further response to drought stimulus (Knight *et al.*, 1998), while it increased the sensitivity of response to low temperature levels (Knight, 2000). Drought pretreatment increases the magnitude of subsequent drought induced  $[Ca^{2+}]_{cyt}$  transients and increases the level of drought inducible calcium regulated gene expression and stress tolerance (Knight *et al.*, 1998).

Calcium plays an important role in mediating plant environment interactions. While plants possess a unique repertoire of Calmodulin (CaM) related proteins

of yet unknown function, the diverse processes in plants seem to be modulated by Calcium/CaM in response to signals from environment (Arazi *et al.*, 1999; Sunkar *et al.*, 2001). Owing to its ubiquitous nature in abiotic stress signaling, calcium signaling can be considered as a potential node at which cross talk can occur. Bryophytes have the ability to maintain the carbohydrate levels during the stress (Robinson *et al.*, 2000). Generally stress causes oxidative damage and it induces marks of oxidative stress. However, the activities of superoxide dismutase, peroxidase and catalase in the increase rapidly during stress period which indicate an inducible defense capability against ROS (Chen *et al.*, 2013). During the stress environments, plants transmit the signal to cellular machinery to activate adaptive responses. The ability of bryophytes to tolerate metal stress is determined by multiple biochemical pathways. Transmembrane transport proteins that mediate ion fluxes play a crucial role in ionic and osmotic homeostasis under stress environments. Defense proteins protect cells from denaturation and degradation, as well as from oxidative damage following exposure to salt stress in bryophytes. ABA and salt stress positively affect the expression of common genes that participate in protection plant cells from injure, and ABA may be responsible for the ability to tolerate stress (Wang *et al.*, 2008).

### 2.2. Calcium sensors

Calcium regulated effector proteins control various processes after calcium elevation. Calcium sensors might also serve as nodes at which cross talks can occur.  $[Ca^{2+}]_{cyt}$  elicited by specific stress could regulate phosphatase and kinase involved in transduction of other stimulus. A variety of calcium sensors have been recognized to play pivotal role in stress responsive signaling.

- CDPKs: Calcium dependent protein kinases. In rice, expression of OsCDPK7 at mRNA level is induced by cold and salt stress (Saijo *et al.*, 2000).
- Calmodulins: Calmodulin has been implicated in plant response to cold, mechanical stimulation, oxidative stress and heavy metal stress (Braam and Davis, 1990; Botella and Arteca, 1994).
- Calcium regulated phosphatases: In *Alfalfa* cells, cold induced inactivation of protein phosphatase 2A (PP2A) is controlled by  $Ca^{2+}$  influx, but PP2A activity could also be modulated by other stress induced  $[Ca^{2+}]_{cyt}$  elevations (Monroy *et al.*, 1998). ABI1 and ABI2 proteins in *Arabidopsis* are potential nodes for cross talks between different signaling pathways involving ABA eg. drought and cold (Sheen, 1998; Sunkar *et al.*, 2001).

### 2.3. MAPK cascades

MAPK cascades are activated by numerous abiotic stresses (Ligterink and Hirt, 2001). Various abiotic stresses rapidly activate *Arabidopsis* MAP kinase ATMPK4 and ATMPK6 providing an evidence for cross talk between MAPK cascades leading separately to ATMPK6 and ATMPK4 (Ichimura *et al.*, 2000).

### 2.4. Transcription factors

Many genes that are induced by cold are also induced by drought or ABA (Shinozaki and Yamaguchi-Shinozaki, 2000), probably because many cold inducible genes encode proteins to protect the plant from the consequences of freezing stress, which includes dehydration. The gene *RD29A* has been utilized in several studies examining the convergence of these pathways (Yamaguchi-Shinozaki and Shinozaki, 1994). In *Arabidopsis*, two groups of transcription factors, DREB1 and DREB2, bind to the cis acting elements (Stockinger *et al.*, 1997; Liu *et al.*, 1998; Shinozaki and Yamaguchi, 2000). The *DREB1* and *DREB2* genes encode structurally different proteins and are induced specifically by low temperature and by salt and drought,

respectively. Overproduction of either DREB1 or DREB2 proteins in protoplast increased expression of an artificial RD29A-promoter GUS fusion gene (Liu *et al.*, 1998), indicating that the DREB promoter element is a point at which drought or salt and cold signal transduction pathways converge and that it can integrate information about these two stimuli.

### 2.5. Reactive oxygen intermediates

Oxidative stress is a common signaling event in all stress situations. Independent of the type of stress influence (natural or anthropogenic), the accumulation of reactive oxygen intermediates is an established fact. Reactive oxygen intermediates (ROIs) are partially reduced forms of atmospheric oxygen. They typically result from excitation of  $O_2$  to form singlet oxygen [ $O_2^1$ ] or from the transfer of one, two or three electrons to  $O_2$  to form, respectively, superoxide radical [ $O_2^-$ ], hydrogen peroxide [ $H_2O_2$ ] or a hydroxyl radical [ $HO^\cdot$ ]. ROIs are capable of unrestricted oxidation of various cellular components and can lead to the oxidative destruction of cellular components (Asada, 1999; Dat *et al.*, 2000). It is well known that water deficit (Sell and Hendry 1992),

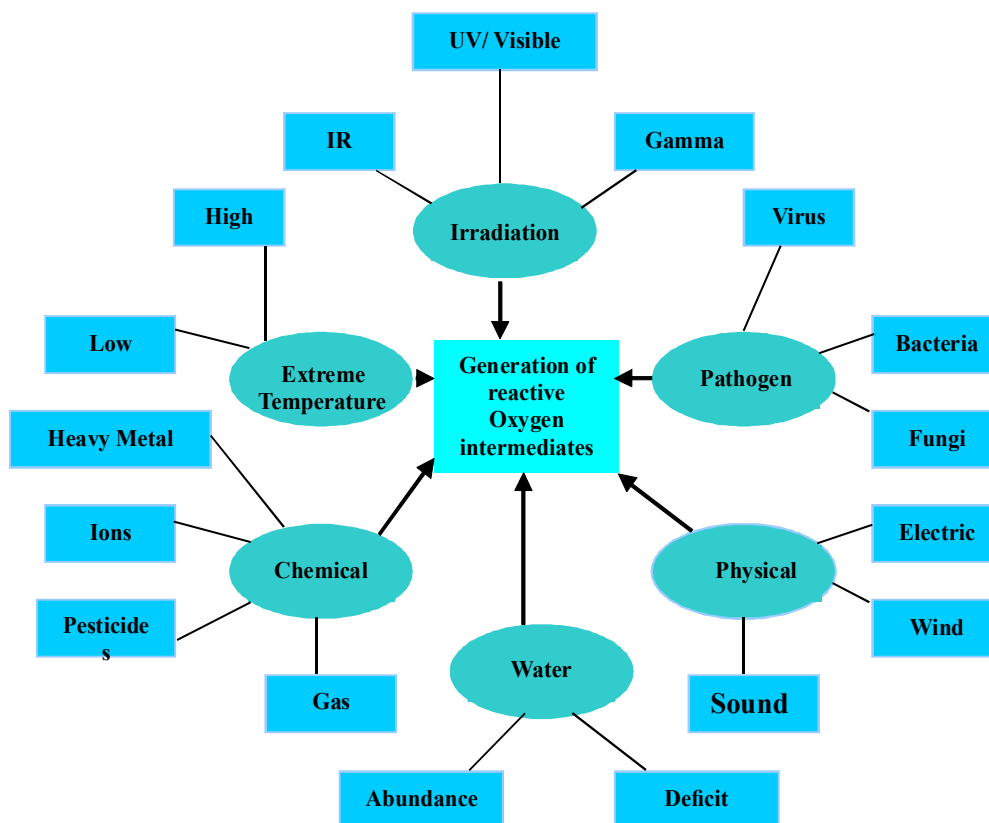


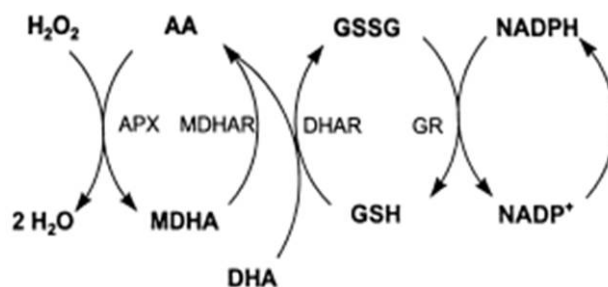
Fig. 1: Agents leading to generation of reactive oxygen intermediates (Modified from Alexieva *et al.*, 2003)

low temperatures (Badiani *et al.*, 1993; Bridger *et al.*, 1994), radiation (Dunning *et al.*, 1994), heavy metals (Gille and Sigler, 1995), acid rains (Velikova *et al.*, 2000), high solar irradiation (Lichtenthaler, 1996) etc. cause an increased production of toxic oxygen species ( $O^2$ ,  $H_2O_2$ ,  $OH$ ,  $O_2^{\cdot-}$ ), which are highly detrimental to all biological systems (Fig. 1). Plants have developed complex antioxidant protective systems in order to cope with all these destructive effects.

Most important common response of plants to different abiotic stresses is accelerated production of reactive oxygen intermediates (ROI). They act as secondary messengers in stress response signal transduction pathways. Abiotic stresses enhance ROI production in chloroplast and mitochondria, while on the other hand by inducing ROI scavenging enzymes reduce ROI levels. ROI production takes place in mitochondria during reduction of respiratory electron transport chain (Moller, 2001) and scavenged by Ascorbate peroxidase (APX) or Superoxide Dismutase (SOD). In peroxisomes, ROIs are produced by glycolate oxidase and  $\beta$ -fatty acid oxidation. Catalase (CAT) is responsible for ROI detoxification during stress, when high level of ROIs is produced (Willekens, 1997). The cytosol with ascorbate glutathione cycle and peroxisomes with CAT act as buffer zones to control overall level of ROIs that reach different cellular compartments during stress and normal metabolism (Mittler, 2002; Mittler *et al.*, 2004). Balance between activities of CAT, SOD and APX in cell is essential for maintaining the steady state levels of ROIs which together with sequestering of metal ions prevent formation of highly toxic hydroxyl radicals via Fenton reaction (Asada and Takahashi, 1987; Willekens, 1997; Asada, 1999).

## 2.6. Redox mediated signaling

$H_2O_2$  acts as a signal-transducing molecule in optimal and stress conditions. This molecule is central to cross tolerance phenomenon and is a key component of stress survival network. Upon elicitation,  $H_2O_2$  is produced in apoplasm by several processes including activation of NADPH oxidase, cell wall peroxidase or other related enzymes i.e. SOD (Bolwell *et al.*, 1995).  $H_2O_2$  has a strong regulatory influence on the fluxes through calcium channels and on calcium concentration in different cellular compartments. The life span of  $H_2O_2$  in a cell depends upon two antioxidant buffers in a cell, namely, ascorbate and glutathione together with antioxidant enzymes that use these antioxidants (Pastori and Foyer, 2002). Ascorbate and glutathione remove  $H_2O_2$  via the Halliwell-Asada pathway (Fig. 2) (Noctor and Foyer, 1998).



**Fig. 2:** Halliwell-Asada pathway or ascorbate glutathione cycle. APX, ascorbate peroxidase; MDHAR, monodehydroascorbate reductase; DHAR, dehydroascorbate reductase; GR, glutathione reductase (May *et al.*, 1998)

Thus it can be concluded that signaling pathways leading to various responses to different stresses constitute a network that is somehow directly or indirectly interconnected at various levels. Various molecules play important roles in helping bryophytes combat a variety of environmental stresses. Bryophytes are simple and appropriate experimental models that enable the researcher to biochemically and genetically dissect out the responses of plants to heavy metal stress, which has become a major area of concern for the environmentalist and the mankind in general (Ingemar *et al.*, 2001; Yayintas *et al.*, 2007).

## 3. Bryophytes as model systems for heavy metal stress tolerance studies

To be successfully employed as a biomonitor in a heavy metal contaminated zone, a plant ought to be fast growing besides having ability to accumulate large quantities of environmentally important metal contaminants. Bryophytes qualify as promising candidates for such studies. Widespread occurrence of these plants and their ability to accumulate metals has lead to their use in environmental monitoring programmes (Wittig, 1993; Conti and Cecchetti, 2001).

Bryophytes facilitate the assessment of overall environmental condition of a given zone (on the basis of stress analysis) alongwith the environmental concentration of a particular contaminant that is present (from tissue analysis). Furthermore, they integrate changes in concentration of metals through time, thereby providing biologically weighted average (Ruhling and Tyler, 1968; Puckett, 1988). Owing to their incredible regeneration capacity, any part of plant can regenerate a protonema in some species, under appropriate conditions. Genetic transformation studies in bryophytes have revealed that integration of foreign DNA sequences in the genome occurs preferentially at targeted location by homologous recombination

(Schaefer, 2002). Thus, the powerful genetic tools can be directly applied for various studies in bryophytes.

Changes in nitrate reductase activity and oxidative stress response in the moss *Polytricum commune* subjected to Chromium, Copper and Zinc phytotoxicity have been reported (Panda and Choudhary, 2005). Induction of oxidative stress and ultrastructural changes are reported in moss *Taxithelium nepalense* under Pb and As phytotoxicity (Choudhary and Panda, 2004). The mechanism of extracellular superoxide radical formation and the role of the oxidative burst in response to desiccation stress is reported in bryophytes and lichens (Minibayeva and Beckett, 2001). Gene structure and expression pattern analysis of three monodehydroascorbate reductase (*Mdhar*) genes in *Physcomitrella patens*, a key enzyme in ascorbate glutathione pathways, playing a major role in detoxification of ROI in plants have been published (Lunde *et al.*, 2006). Metabolism of Reactive oxygen intermediates in desiccation-stressed thalli of the liverwort *Dumortiera hirsuta* has been studied (Beckett *et al.*, 2004). Mayaba *et al.* (2002) reported an oxidative burst of hydrogen peroxide during rehydration following desiccation in the moss *Atrichum androgynum*.

#### 4. Various possible targets for studying heavy metal stress tolerance in Bryophytes

##### 4.1. Micro RNAs

Micro RNAs (miRNA) are one class of endogenous tiny (21-22 nucleotide) non-coding RNA that play important regulatory roles in plant development and response to external stimuli (Bartel and Bartel, 2003; Kidner and Martienssen, 2005). miRNA controls gene expression post transcriptionally by targeting cognate mRNAs for degradation (Llave *et al.*, 2002; Palatnik *et al.*, 2003) or by translational repression (Reinhart *et al.*, 2000; Aukerman and Sakai, 2003; Chen, 2004). To identify miRNAs, a library of small RNAs from protonema of *Physcomitrella patens* was constructed and several distinct sRNA expression patterns were observed (Arazi *et al.*, 2005). MiR156, Pp-miR319b, miR390, Pp\_71 and Pp\_89 were expressed at similar levels in the juvenile as well as adult phases of the gametophyte. Higher expression levels of Pp\_42 and Pp\_94 were observed in *P. patens* differentiating caulonema cells, while Pp\_60 was detected almost entirely in non-differentiated protonema that is mainly composed of chloronema cells. It can be speculated that Pp\_60 might function in an opposite way to Pp\_94 and downregulates one of the factors that regulates chloronema differentiation (Arazi *et al.*, 2005). The transition from chloronema cells to caulonema is an

indication of stress in bryophytes and thus miRNAs might serve as promising tools for stress tolerance studies in bryophytes (Schween *et al.*, 2003).

##### 4.2. ALDH11A5 : a novel non-phosphorylating GAPDH cDNA

A cDNA ALDH11A5 (AY504666) was identified in *P. patens* with significant similarity to cytosolic NADP<sup>+</sup>-dependent, non-phosphorylating glyceraldehyde-3-phosphate dehydrogenase (GAPN; EC 1.2.1.9) (Wood *et al.*, 2004). GAPN is member of ALDH protein subfamily (Wood and Kravesky, 2002). ALDH11 (GAPN) orthologues operate in cytosol where enzymes generate NADPH for biosynthetic processes (Gao and Loescher, 2000). Increased GAPN activity allows glycolysis to continue under stress conditions that limit adenylate nucleotide concentration. GAPN glycolytic shunt continues to function in response to environmental stress (Plaxton, 1996). The expression level of ALDH11A5 was studied in response to osmotic stress, salt stress and ABA (Wood *et al.*, 2004). Further molecular and physiological experiments can evaluate the role of ALDH11A5 in plant development and adaptation to environmental stresses.

##### 4.3. Sugar: 3-O-Methyl rhamnose

Primary cell wall (PCW) plays an indispensable role during stress tolerance in plant systems. One major wall related difference between bryophytes and higher plants is that bryophytes do not have lignified conducting tissues. The major sugar residues in PCW of bryophytes are galactose, mannose, xylose, arabinogalactan, rhamnose etc (Thomas, 1977). Although monosaccharides present in bryophytes are similar to those found in angiosperms, there are quantitative differences between PCW composition between bryophytes and angiosperms: bryophyte PCWs contain more galactose A, galactose C and mannose than those of angiosperms (Popper and Fry, 2003). Immunocytochemical studies of bryophyte cell walls indicate that they contain high concentrations of rhamnogalacturonan-I, homogalacturonan and arabinogalactan proteins (Ligrone *et al.*, 2002). 3-O-Methylrhamnose found in high concentrations particularly in bryophyte's primary cell walls may be associated with an acidic polysaccharide which helps in preventing dessication in bryophytes (Popper *et al.*, 2004).

##### 4.4. IQD1: Calmodulin binding nuclear proteins

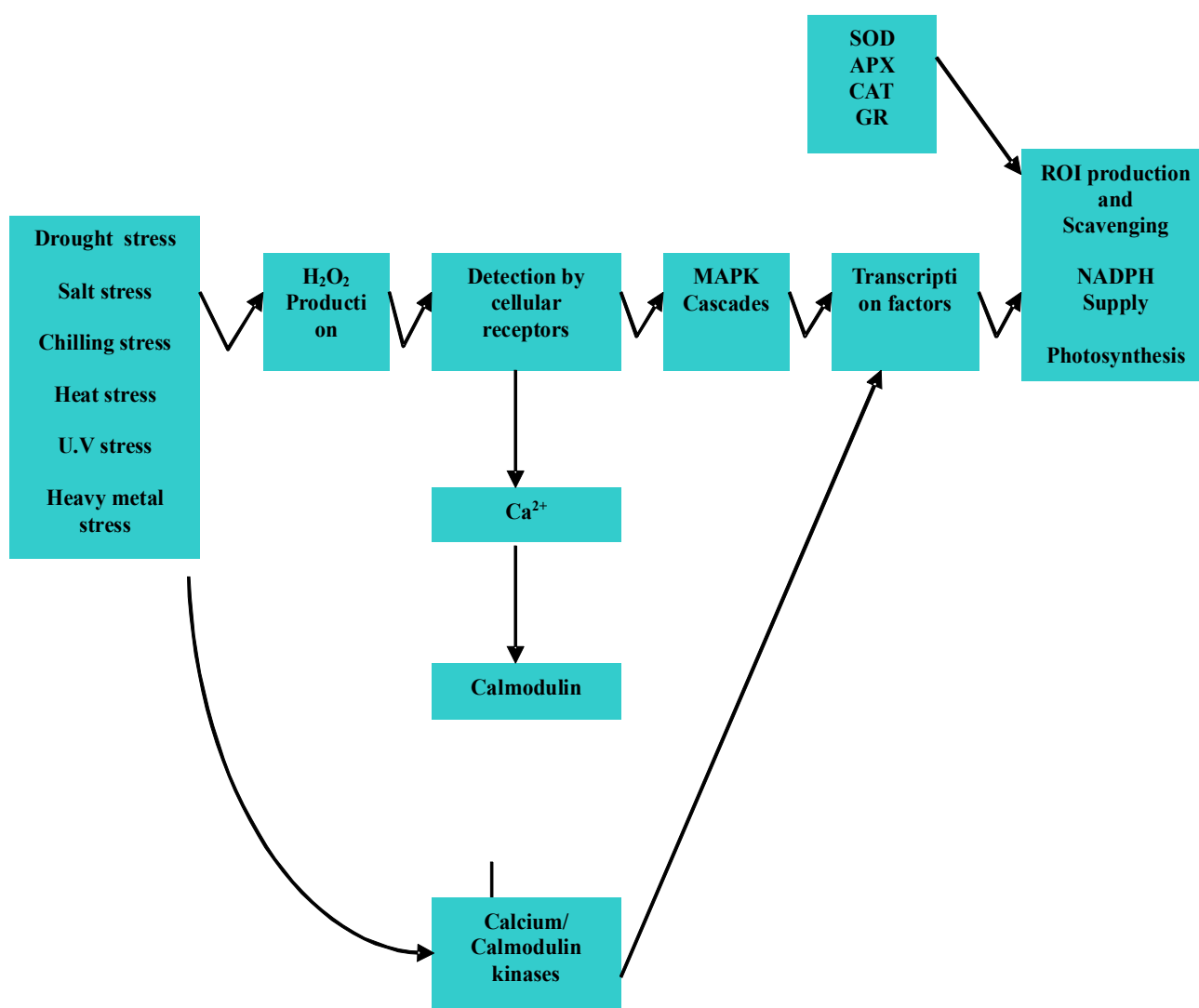
IQD1 are novel calmodulin binding nuclear proteins, which are known to stimulate glucosinolate and plant defense in *Arabidopsis thaliana*. One contig EST sequence from *Physcomitrella patens*, IQD like protein

(contig 5180) has been identified (Abel *et al.*, 2005), which can be a potential target for heavy metal stress tolerance (Fig 3). Although the deduced amino acid sequence appears to be truncated at the C-terminus (20 amino acid residue downstream of IQ67 domain) an appreciable similarity to the protein encoded by At1g01110 is found (33% identity) which includes the presence of MEME motif 3 at its N-terminus. Alignment of IQ67 domain of moss polypeptide reveals a deletion of six residues that correspond to the N-terminus of the second IQ67 domain encoding exon of most *Arabidopsis* and *Rice* IQD proteins. As the IQ67 intron is in phase-0 and since both *A. thaliana* and *Oryza sativa* express an IQD-like gene in which the second IQ67 domain-encoding exon is replaced by an unrelated exon, it is

unlikely that the contig 5180 DNA sequence is an artifact and probably represents either a novel variant of IQD-like genes or an ancestral gene of the IQD genes found in vascular plants (Abel *et al.*, 2005).

#### 4.5. Calmodulin binding proteins in bryophytes

Plant response to a variety of environmental stresses is mediated in part by signaling processes involving cytosolic calcium and calcium binding protein, calmodulin (Knight *et al.*, 1997; Plieth *et al.*, 1999; Sanders *et al.*, 1999). Genes encoding novel membrane transporter like proteins– Mcamb1, Mcamb2 have been identified in *Physcomitrella patens* (Takezawa and Minami, 2004). Calmodulin binds to Mcamb1 and Mcamb2 via interaction with basic



**Fig. 3:** Activation of signal transduction events during oxidative stress exhibiting potential nodes where cross talks can occur (Modified from Mittler, 2002)

amphiphilic amino acids in C-terminal domains. Levels of *Mcamb1* and *Mcamb2* transcripts increase dramatically following treatments with low temperature, hyperosmotic solute and stress hormone ABA. Calmodulin participates in cellular signaling events leading to increase in stress resistance through regulation of novel transporter like proteins (Takezawa and Minami, 2004). Hence, studies indicate that calcium might play a role in signaling events leading to the development of stress resistance through modulation of stress inducible CaM-binding transporters in bryophytes (Fig 3).

#### 4.6. GC-rich repetitive DNA sequences

Behaviour of repetitive DNA in response to lead and cadmium in two bryophytes namely *Leptodictyum riparium* and *Funaria hygrometrica* was studied (Bassi *et al.*, 2006). Metal treatment was accompanied by selective amplification of some GC-rich repetitive DNA sequences forming peculiar agglomerates inside the nucleus (Bassi *et al.*, 1995; Bassi, 1999). This amplification is quantitatively proportional to the time of exposure of the plants to metals and stops upon removal of metals from the culture medium. Results show that ribosomal DNA sequences are involved in this metal-induced repetitive DNA agglomerates formation (Bassi *et al.*, 2006). Thus focus should be on this particular fraction of the genome to identify the potential targets for stress tolerance in bryophytes.

#### 4.7. Expansins

Expansins are extracellular proteins involved in cell wall loosening (Cosgrove, 2000). They play a definite role in plant growth and differentiation (caulonema formation). Expansins like proteins are reported in *Physcomitrella patens* (Schipper *et al.*, 2002). Both alpha ( $\alpha$ ) and beta ( $\beta$ ) expansins are present as gene families in this plant and expression analysis indicates that these genes respond to a complex regulation by hormonal and environmental factors. In particular, the expression of many expansins genes gets upregulated in stress conditions suggesting that they play a definite role in specific cellular differentiation displayed by *Physcomitrella patens* in response to such stresses. Six cDNA encoding expansins like proteins are found in *Physcomitrella patens*. PpExp1, PpExp2, PpExp and PpExp4 encode  $\alpha$ -expansins like proteins, whereas PpExpB1 and PpExpB2 encode  $\beta$ -expansins like proteins (Schipper *et al.*, 2002).

These provide a powerful tool to investigate the purported role of expansins in modulating plant growth and form via cell wall extensibility in response to stress conditions.

#### 4.8. Messenger Ribonucleoprotein particles (mRNPs)

Alteration in gene expression elicited by stress is primarily regulated at the translational level (Oliver, 1991; Scott and Oliver, 1994; Oliver and Wood, 1997). The alteration in protein synthesis patterns during rehydration occurs as a result of differential selection of rehydrin mRNAs from a constant mRNA pool. mRNPs are proteins that permit restoration of protein synthesis following rehydration (Wood and Oliver, 1999). The studies on *Tortula ruralis* indicate that in mosses, the selection of rehydrin mRNA is in part aided by storage of these transcripts as messenger ribonucleoprotein particles (mRNPs) during drying.

Tr288 is a rehydrin in *Tortula ruralis* that codes for a protein having structural similarity to higher plants LEA proteins (Wood and Oliver, 1999). This rehydrin has a transcript that is abundant during slow drying and accumulates in polysomal fractions of cell extracts from slow dried gametophytes. The polysomal retention of Rps3a and Rps14, Rps16 and Rp123 transcripts in desiccated moss gametophyte indicates that not all transcripts made in response to a stress event are required for immediate use but may be synthesized and stored for the recovery period (Wood *et al.*, 2000). The formation of mRNPs in response to water loss and their possible roles in mRNA storage and protection has important consequences for the study of vegetative desiccation tolerance and perhaps stress responses of plants in general.

The ability to store components during any stress event that are needed for recovery offers a new dimension to the concept of damage control and rapid return to growth than does relatively slower activation and transcription of specific stress recovery genes thus making mRNPs promising tools for stress tolerance in bryophytes (Wood and Oliver, 1999).

Various field and culture studies have been carried out using bryophytes as heavy metal monitoring tools (Kuik and Wolterbeek, 1995; Markert *et al.*, 1996; Sucharova and Suchara 1998; Bargagli *et al.*, 2002; Schroder and Pesch 2005; Tyagi *et al.*, 2007). But, information on the relationship between heavy metal stress on bryophytes at biochemical, protein and gene level is rather incomplete.

#### 5. Recommendations

Heavy metal pollution is a fast growing environmental problem, which calls for immediate attention. Owing to the higher cost outlay, inability to detect short lived deleterious effects and minute

quantities of pollutants below the detection limits of conventional remediation technologies, the need of the hour is to use plants for environmental cleanup. Bryophytes qualify as promising candidates for such studies as they need minimum care during their life cycle. Surprisingly, potential of these plants has always been underestimated. Heterotrimeric guanine nucleotide binding proteins (G-proteins) are well characterized signaling molecules that interact with plasma membrane localized G-protein coupled receptors (GPCRs) and transduce majority (~80%) of extracellular signals across the cell membranes. However, these functions are believed to be performed by regulator of G-protein signaling (RGS) in plants. The main objective of this review is to provide description of the relevant molecules that might play leading roles in heavy metal stress tolerance in bryophytes. There are several processes including signal transduction, transcript directive, homeostasis and biosynthesis defense proteins which need to be thoroughly addressed to understand the stress tolerance mechanism in bryophytes. We, hope to promote the exploration of bryophytes at biochemical, protein and gene level as tools for combating heavy metal pollution.

## References

- Abel, S., Savchenko, T., and Levy, M. 2005. Genome wide comparative analysis of the I Q D gene families in *Arabidopsis* and *Oryza sativa*. *BMC Evolutionary Biology* **5**:72-87.
- Alexieva, V., Ivanov, S., Sergiev, I. and Karanov, E. 2003. Interaction between stresses. *Bulgarian Journal of Plant Physiology* (special issue) pp. 1-17.
- Angelone, M. and Bini, C. 1992. Trace elements concentrations in the soils and plants of western Europe. In: Adriano, D.C. (Ed.). *Biochemistry of Trace Elements*. FL: Lewis Publishers, Boca Raton, pp. 19-60.
- Arazi, T., Neiman, M.T., Stav, R., Riese, M., Hujiser, P. and Baulcome, D.C. 2005. Cloning and characterization of micro-RNAs from mosses. *Plant Journal* **43**:837-848.
- Arazi, T., Sunkar, R., Kaplan, B. and Fromm, H. 1999. A tobacco plasma membrane calmodulin binding transporter confers  $\text{Ni}^{2+}$  tolerance  $\text{Pb}^{2+}$  hypersensitivity in transgenics plants. *Plant Journal* **20**:171-182.
- Asada, K. 1999. The water-water cycle in chloroplasts: scavenging of active oxygen and dissipation of excess photons. *Annual Review of Plant Physiology and Plant Molecular Biology* **50**:601-639.
- Asada, K. and Takahashi, M. 1987. Production and scavenging of active oxygen in photosynthesis. In: Kyle D.J., Osmond C.B. and Arntzen C.J. (Eds.). *Photoinhibition*. Elsevier, Amsterdam, pp. 227-287.
- Aukerman, M.J. and Sakai, H. 2003. Regulation of flowering time and floral organ identity by a microRNA and its APETALA2-like targets genes. *Plant Cell* **17**:2730-2741.
- Badiani, M.A., Paolacci, A., D'Annibale, G. and Sermanini 1993. Antioxidant and photosynthesis in the leaves of *Triticum durum* L. seedlings acclimated to low, non-chilling temperature. *Journal of Plant Physiology* **142**:18-24.
- Bargagli, R., Monaci, F., Borghini, F., Bravi, F. and Agnorelli C. 2002. Mosses and lichens as biomonitors of trace metals. A comparative study on *Hypnum cupressiforme* and *Parmelia caperata* in a former mining district in Italy. *Environmental Pollution* **116**:279-287.
- Bartel, B. and Bartel, D.P. 2003. Micro RNAs: at the root of plant development? *Plant Physiology* **132**:709-717.
- Bassi, P. 1999. The effect of environmental stress on the repetitive DNA behaviour in plants. In: Lerner H. R. (Ed.). *Plant responses to Environmental Stresses*. Marcel Dekker, New York, pp. 161-170.
- Bassi, P., Basile, A., Ferraro, M., Masi, M., Migliaccio, D., Morelli, G. and Napolitano, E. 2006. Plasticity of repetitive DNA in response to metal stress in bryophytes. *Plant Biosystems* **140**:80-86.
- Bassi, P., Basile, A., Stefanini, A., Vosa, C.G., Maffei, M.A., Giordano, S. and Castaldo, R. 1995. Effects of lead on the nuclear repetitive DNA of the moss *Funaria hygrometrica*. *Protoplasma* **188**:104-108.
- Beckett, R.P., Farida, V., Minibayeva, F., Luthje, S. and Bottger, M. 2004. Reactive oxygen species metabolism in desiccation-stressed thalli of the liverwort *Dumortiera hirsuta*. *Physiologia Plantarum* **122**:3-10.
- Bolwell, G.P., Buti, V.S., Davies, D.R. and Zimmerlin, A. 1995. The origin of the oxidative burst in plants. *Free Radical Research Communications* **23**:517-532.
- Botella, J.R. and Arteca, R.N. 1994. Differential expression of two calmodulin genes in response to physical and chemical stimuli. *Plant Molecular Biology* **24**:757-766.
- Braam, J. and Davis, R.W. 1990. Rain, wind and touch induced expression of calmodulin and calmodulin related genes in *Arabidopsis*. *Cell* **60**:357-364.
- Breckle, C.W. 1991. Growth under heavy metals. In: Waisel Y., Eshel A., and Kafafi U. (Eds.). *Plant Roots: The Hidden Half*. Marcel Dekker, New York, pp. 351-375.
- Bridger, G.W., Yang, D., Falk, B. and Kerstie, Mc 1994. Cold acclimation increased tolerance of activated oxygen in winter cereals. *Journal of Plant Physiology* **144**:235-240.
- Chaudhary, S. and Panda, S.K. 2004. Induction of oxidative stress and ultrastructural changes in moss *Taxithelium nepalense* under lead and arsenic phytotoxicity. *Current Science* **87**:342-347.
- Chen, W.J., Zhang, N., Hang, L.L., Wang, Y., Ji, Mc 2013. Influence of shading during the processes of drought stress and re-watering on the physiological and biochemical characteristics of *Haplocladium microphyllum*. *Chinese Journal of Applied Ecology* **24**:57-62.
- Chen, X. 2004. A micro RNA as a translator repressor of APETALA2 in *Arabidopsis* flower development. *Science* **303**:2022-2025.
- Chrestensen, C.A., Starke, D.W. and Mieyal, J.J. 2000. Acute Cadmium exposure inactivates thioltransferase



- (glutaredoxin), inhibits intracellular reduction of proteins-glutathionyl-mixed disulphides, and initiates apoptosis. *The Journal of Biological Chemistry* **275**:26556-26565.
- Conti, M.E. and Cecchetti, G. 2001. Biological monitoring: lichens as bioindicators of air pollution assessment – a review. *Environmental Pollution* **114**:471-492.
- Cosgrove, D.J. 2000. Loosening of plant cell walls by expansins. *Nature* **407**:321-326.
- Dat, J., Vandenabeele, S., Vranová, E., Van Montagu, M., Inzé, D. and Van Breusegem, F. 2000. Dual action of the active oxygen species during plant stress response. *Cell and Molecular Life Sciences* **57**:779-795.
- Dunning, C.L., Chalker, S.J. and Scott, A. 1994. Exposure to UV-B radiation increased cold hardness in *Phododendron*. *Plant Physiology* **92**:516-520.
- Galloway, J.N., Thornton, J.D., Norton, S.A., Volcho, H.L. and Mc Lean, R.A.N. 1982. Trace metals in atmospheric deposition: a review and assessment. *Atmospheric Environment* **16**:1677-1700.
- Gao, Z. and Loescher, W.H. 2000. NADPH supply and mannitol biosynthesis, characterization, cloning and regulation of the non- reversible glyceraldehydes-3- phosphate dehydrogenase in celery leaves. *Plant Physiology* **124**:321-330.
- Gille, G. and Sigler, K. 1995. Oxidative stress and living cells. *Folia Microbiology* **40**:131-152.
- Godbold, D.L. and Huttermann, A. 1985. Effect of zinc, cadmium and mercury on root elongation of *Picea abies* (Karst.) seedlings and the significance of these metals to forest dieback. *Environmental Pollution* **38**:375-381.
- Ichimura, K., Mizoguchi, T., Yoshida, R., Yuasa, T. and Shinozaki, K. 2000. Various abiotic stresses rapidly activate *Arabidopsis* MAP kinases ATMPK 4 and ATMPK6. *Plant Journal* **24**:655-665.
- Imlay, A., Chin, S.M. and Linn, S. 1988. Toxic DNA damage by hydrogen peroxide through the Fenton reaction *in vivo* and *in vitro*. *Science* **240**:640-642.
- Ingemar, R., Richard, B., Emily, B., Ove, E. and Suzanne, M. 2001. Sediment Evidence of Early Eutrophication and Heavy Metal Pollution of Lake Mälaren, Central Sweden. *AMBIO: A Journal of the Human Environment* **30**:496-502.
- Jones, P., Kortenkamp, A., O'Brien, P., Wang, G. and Yang, G. 1991. Evidence for the generation of hydroxyl radicals from a chromium (V) intermediate isolated from the reaction of chromate with glutathione. *Biochimica et Biophysica Acta* **286**:652-655.
- Kidner, C.A. and Martienssen, R. A. 2005. The developmental role of microRNA in plants. *Current Opinions in Plant Biology* **8**:38-44.
- Knight, H. 2000. Calcium signaling during abiotic stress in plants. *International Review of Cytology* **195**:269-324.
- Knight, H. and Knight, M.R. 2001. Abiotic stress signaling pathways: specificity and cross talks. *Trends in Plant Science* **6**:262-267.
- Knight, H., Brandt, S. and Knight, M.C. 1998. A history of stress alters drought calcium signaling pathways in *Arabidopsis*. *Plant Journal* **16**:681-687.
- Knight, H., Trewavas, A.J. and Knight, M.R. 1997. Calcium signaling in *Arabidopsis thaliana* responding to drought and salinity. *Plant Journal* **12**:1067-1078.
- Kuik, P. and Wolterbeek, H.T. 1995. Factor analysis of atmospheric trace-element deposition data in the Netherlands obtained by moss monitoring. *Water, Air and Soil Pollution* **84**:323-346.
- Lanzty, R.J. and Mackenzie, F.T. 1979. Atmospheric trace metals: global cycles and assessment of man's impact. *Geochimica et Cosmochemica Acta* **43**:511-525.
- Lichtenthaler, H.K. 1996. Vegetation stress: an introduction to the stress concept in plants. *Journal of Plant Physiology* **148**:4-14.
- Ligrone, R., Vaughn, K.C., Renzaglia, K.S., Knox, J.P. and Duckett, J.G. 2002. Diversity in the distribution of polysaccharides and glycoprotein epitopes in the cell walls of bryophytes: new evidence for the multiple evolution of water conducting cells. *New Phytologist* **156**:491-508.
- Ligterink, W. and Hirt, H. 2001. Mitogen activated protein (MAP) kinase pathways in plants: versatile signaling tools. *International Review of Cytology* **201**:209-275.
- Liu, Q., Kasuga, M., Sakuma, M., Abe, H., Miura, S., Yamaguchi-Shinozaki, K. and Shinozaki, K. 1998. Two transcription factors, DREB1 and DREB2, with an EREBP/AP2 DNA binding domain separate two cellular signal transduction pathways in drought and low temperature responsive gene expression respectively in *Arabidopsis*. *Plant Cell* **10**:1391-1406.
- Llave, C., Xie, Z., Kasschau, K.D. and Carrington, J.C. 2002. Cleavage of scare crow like mRNA targets directed by a class of *Arabidopsis* miRNA. *Science* **297**:2053-2056.
- Lund, B.O., Miller, D.M. and Woods, J.S. 1991. Mercury induced H<sub>2</sub>O<sub>2</sub> production and lipid peroxidation *in vitro* in rat kidney mitochondria. *Biochemical Pharmacology* **42**:181-187.
- Lunde, C., Baumann, U., Shirley, J., Drew, D.P. and Fincher, G.B. 2006. Gene structure and expression pattern analysis of three monodehydroascorbate reductase (*Mdhar*) genes in *Physcomitrella patens*: implications for the evolution of the MDHAR family in plants. *Plant Molecular Biology* **60**:259-275.
- Markert, B., Herpin, U., Berlekamp, J., Oehlmann, J., Grodzi, N'ska, K., Mankovska, B., Suchara, I., Siewers, U., Weckert, V. and Lieth, H. 1996. A comparison of heavy metal deposition in selected Eastern European countries using the moss monitoring method, with special emphasis on the "Black Triangle". *The Science of the Total Environment* **193**:85-100.
- May, M.J., Vernoux, T., Leaver, C., Van Montagu, M. and Inze, D. 1998. Glutathione homeostasis in plants: implications for environmental sensing and plant development. *Journal of Experimental Botany* **49**:649-667.
- Mayaba, N., Minibayeva, F. and Beckett, R.P. 2002. An oxidative

- burst of hydrogen peroxide during rehydration following desiccation in the moss *Atrichum androgynum*. *New Phytologist* **155**:275-282.
- Minibayeva, F. and Beckett, R.P. 2001. High rates of extracellular superoxide production in bryophytes and lichens, and an oxidative burst in response to rehydration following desiccation. *New Phytologist* **152**:333-340.
- Mittler, R. 2002. Oxidative stress, antioxidants and stress tolerance. *Trends in Plant Science* **7**:405-410.
- Mittler, R., Vanderauwera, S., Gollery, M. and Breusegem, F.V. 2004. Reactive oxygen gene network of plants. *Trends in Plant Science* **9**:490-498.
- Moller, I.M. 2001. Plant mitochondria and oxidative: electron transport, NADPH turnover, and metabolism of reactive Oxygen species. *Annual Review of Plant Physiology and Plant Molecular Biology* **52**:561-591.
- Monroy, A.F., Sangwan, V. and Dhindsa, R. (1998). Low temperature signal transduction during cold acclimation: Protein phosphatase 2A as an early target for cold inactivation. *Plant Journal* **13**:653-660.
- Moriarty, F. 1999. Ecotoxicology-The Study of Pollutants. In: Moriarty F. (Ed.) *Ecosystems*. Academic Press. San Diego, pp.347.
- Nies, D.H. 1999. Microbial heavy metal resistance. *Applied Microbiology and Biotechnology* **51**:730-750.
- Noctor, G. and Foyer, C. 1998. Ascorbate and glutathione : keeping active oxygen under control . *Annual Review of Plant Physiology and Molecular Biology* **49**:249-279.
- Nriagu, J.O. 1996. History of global metal pollution. *Science* **272**:223.
- Oliver, M.J. 1991. Influence of the protoplasmic water loss on the control of protein synthesis in the dessication tolerant moss *Tortula ruralis*: Ramifications for a repair based mechanism of dessication tolerance. *Plant Physiology* **97**:1501-1511.
- Oliver, M.J. and Wood, A.J. 1997. Desiccation tolerance in mosses. In: T. Koval (Ed.). *Stress-Inducible Processes in Higher Eukaryotic Cells*. Plenum Press, New York, pp. 1-26.
- Oliver, M.J., Velten, J. and Mishler, B.D. 2005. Desiccation Tolerance in Bryophytes: A Reflection of the Primitive Strategy for Plant Survival in Dehydrating Habitats? *Integrative and Comparative Biology* **45**:788-799.
- Palatnik, J.F., Allen, E., Wu, X., Schommer, C., Schwab, R., Carrington, J.C. and Weigel, D. 2003. Control of leaf morphogenesis by micro RNAs. *Nature* **425**:257-263.
- Panda, S.K. and Chaudhary, S. 2005. Changes in nitrate reductase activity and oxidative stress response in the moss *Polytrichum commune* subjected to chromium, copper and zinc phytotoxicity. *Brazilian Journal of Plant Physiology* **17**:25-27.
- Pastori, G.M. and Foyer, C.H. 2002. Common component networks, and pathways of cross-tolerance to stress. The central role of "Redox" and abscisic acid-mediated controls. *Plant Physiology* **129**:460-468.
- Plaxton, W. 1996. The organization and regulation of plant glycolysis. *Annual Review of Plant Physiology and Plant Molecular Biology* **47**:185-214.
- Plieth, C., Hansen, H., Knight, H. and Knight, M.R. 1999. The primary characteristics of signal perception and calcium response. *Plant Journal* **18**:491-497.
- Popper, Z.A. and Fry, S.C. 2003. Primary cell wall composition of bryophytes and charophytes. *Annals Botany* **91**:1-2.
- Popper, Z.A., Sadler, I.H. and Fry, S.C. 2004. 3-O-Methylrhamnose in lower land plant primary cell walls. *Biochemical Systematics and Ecology* **32**:279-289.
- Puckett, K.J. 1988. Bryophytes and lichens as monitors of metal deposition. *Bibliotheca Lichenologica* **30**:231-267.
- Reinhart, B.J., Slack, F.J., Bassoon, M., Pasquinelli, A.E., Bettinger, J.C., Rougvie, A.E., Horvitz, H.R. and Ruvkun, G. 2000. The 21 nucleotide let-7 RNA regulates development timing in *Caenorhabditis elegans*. *Nature* **403**:901-906.
- Rivetta, A., Negrini, N. and Cocucci, M. 1997. Involvement of Ca<sup>2+</sup>/calmodulin in Cd<sup>2+</sup> toxicity during the early phases of radish seed germination. *Plant Cell and Environment* **20**:600-608.
- Ruhling, A. and Tyler, G. 1968. An ecological approach to the lead problem. *Botaniska Notiser* **122**:248-342.
- Saijo, Y., Hata S., Kyoizuka, J., Shimamoto, K. and Azui, K. 2000. Overexpression of a single Ca<sup>2+</sup> dependent protein kinase confers both cold and salt/ drought tolerance on rice plants. *Plant Journal* **23**:319-327.
- Sanders, D., Brownlee, J. and Harper, J. 1999. Communicating with Calcium. *Plant Cell* **11**: 691-706.
- Schaefer, D. 2002. A new moss genetics: Targeted Mutagenesis in *Physcomitrella patens*. *Annual Review of Plant Biology* **53**:477-501.
- Schipper, O., Schaefer, D., Reski, R. and Fleming, A. 2002. Expansins in the bryophyte *Physcomitrella Patens*. *Plant Molecular Biology* **50**:789-802.
- Schroder, W. and Pesch, R. 2005. Time series of metals in mosses and their correlation with selected sampling site-specific and ecoregional characteristics in Germany. *Environmental Science and Pollution Research* **12**:159-67.
- Schween, G., Gorr, G. Hohe, A. and Reski, R. 2003. Unique tissue specific cell cycle in *Physcomitrella*. *Plant Biology* **5**:1-9.
- Scott, H.B.I.I. and Oliver, M.J. 1994. Accumulation and polysomal recruitment of the transcripts in response to dessication and rehydration of the moss *Tortula ruralis*. *Journal of Experimental Botany* **45**:577-583.
- Sell, W.G. and Hendry, J.L. 1992. Effect of dessication of some activated oxygen processing enzymes and anti-oxidants in mosses. *Journal of Experimental Botany* **43**:1031-1037.
- Sheen, J. 1998. Mutational analysis of protein phosphatases 2 C involved in abscisic acid signal transduction in higher plants. *Proceedings of National Academy of Sciences USA* **95**:975-980.

- Shi, X., Dalal, N.S. and Kasprzak, K.S. 1993. Generation of free radicals from hydrogen peroxide and lipid hydroperoxides in the presence of Cr (III). *Biochimica et Biophysica Acta* **302**:294-299.
- Shinozaki, K. and Yamaguchi-Shinozaki, K. 2000. Molecular responses to dehydration and low temperature: differences and cross talk between two stresses signaling pathways. *Current Opinions in Plant Biology* **3**:217-223.
- Stockinger, E., Gilmour, S.J. and Thomashow, M.F. 1997. Arabidopsis thaliana CBF1 encodes an AP2 domain containing transcriptional activator that binds to C-repeat/ DRE, a cis acting DNA regulatory element that stimulates transcription in response to low temperature and water deficit. *Proceedings of National Academy of Sciences USA* **94**:1035-1040.
- Sucharova, J. and Suchara, I. 1998. Atmospheric deposition levels of chosen elements in the Czech Republic determined in the framework of the International Bryomonitoring Program 1995. *The Science of the Total Environment* **223**:37-52.
- Sunkar, R., Arazi, T., Kaplan, B., Dolev, D. and Fromm, H. 2001. Calmodulin and plant responses to the environment: modulation of plant tolerance to toxic metals by a plasma membrane calcium/calmodulin binding channel. In: Sopory, S.K., Oelmüller, R and Maheshwari S.C. (Eds.) Kluwer Academic/Plenum Publishers, New York, pp.350.
- Takezawa, D. and Minami, A. 2004. Calmodulin-binding proteins in bryophytes: Identification of abscisic acid, cold and osmotic stress induced gene encoding novel membrane bound transporter like proteins. *Biochemical and Biophysical Research Communication* **317**:428-436.
- Thomas, R.J. 1977. Wall analysis of *Lophocoela* seta cells (bryophyta) before and after elongation. *Plant Physiology* **59**:337-340.
- Tyagi, R., Gupta, P. and Uniyal, P.L. 2007. Studies on effect of Lead and Zinc concentration on growth of four species of bryophytes. *International Journal of Biological and Chemical sciences* **1**:128-135.
- Van Assche, F. and Clijsters, H. 1986. Inhibition of photosynthesis in *Phaseolus vulgaris* by treatment with toxic concentration of Zinc: effect on ribulose-1, 5-biphosphate carboxylase/oxygenase. *Journal of Plant Physiology* **125**:355-360.
- Velikova, V.I., Yordanov, A. and Edreva, A. 2000. Oxidative stress and some antioxidant systems in acid rain treated bean plants. Protective role of exogenous polyamines. *Plant Science* **151**:59-66.
- Wang, X., Liu, Z. and He, Y. 2008. Responses and tolerance to salt. *Plant Signaling Behavior* **3**:516-518.
- Willekens, H., Chamnongpol, S., Davey, M., Schraudner, M., Langebartels, C., Montagu, M.V., Inzé, D. and Van Camp, W. 1997. Catalase is a sink for H<sub>2</sub>O<sub>2</sub> and is indispensable for stress defense in C3 plants. *EMBO Journal* **16**:4806-4816.
- Williams, L.E., Pittman, J.K. and Hall, J.L. 2000. Emerging mechanisms of heavy metal transport in plants. *Biochimica et Biophysica Acta* **1465**:104-126.
- Wittig, R. 1993. General aspects of biomonitoring heavy metals by plants. In: Markert B. (Ed.). *Plants as Biomonitor-Indicators for Heavy Metals in the Terrestrial Environment*. V.H.C. Press, Weinheim, pp. 3-27.
- Wood, A.J., Band Kravesky, S. 2002. The role of aldehyde dehydrogenase (ALDHs) in plant stress tolerance. In: Wood A.J. (Ed.). *Biochemical and Molecular Responses of the Plants to the Environment*. Research Signposts, Trivandrum, pp.1-14.
- Wood, A.J. and Oliver, M.J. 1999. Translational control in plants stress: the formation of messenger ribonucleoprotein particles (mRNPs) in response to desiccation of *Tortula ruralis* gametophytes. *The Plant Journal* **18**:359-370.
- Wood, A.J., Duff, R.J. and Oliver, M.J. 2000. The translational apparatus of *Tortula ruralis*: polysomal retention of transcripts encoding the ribosomal proteins RPS14, RPS16 and RPL23 in desiccated and rehydrated gametophytes. *Journal of Experimental Botany* **51**:1655-1662.
- Wood, A.J., Reski, R. and Frank, W. 2004. Isolation and characterization of ALDH11A5, a novel non-phosphorylating GADPH cDNA from *Physcomitrella patens*. *The Bryologist* **107**:385-387.
- Yamaguchi-Shinozaki, K. and Shinozaki, K. 1994. A novel cis acting element in an Arabidopsis gene is involved in responsiveness to drought, low temperature, or high salt stress. *Plant Cell* **6**:251-264.
- Yayintas, O., Yilmaz, S., Turkoglu, M. and Yusuf, D. 2007. Determination of heavy metal pollution with environmental physicochemical parameters in waste water of Kocabas Stream (Biga, Canakkale, Turkey) by ICP-AES. In: Filar J.A. (Ed.). *Environmental Monitoring and Assessment*. Springer, London, pp. 389-397.